In vivo studies regarding the antioxidant effect of certain vegetal extracts on the stimulation of the antioxidant system in *Oncorhynchus Mykiss*, in conditions of overpopulation

Received for publication, november, 15, 2013. Accepted, december, 1, 2013

Mariana LUPOAE¹, Dragomir COPREAN², Paul LUPOAE³

- ¹ "Dunărea de Jos" University of Galați, Faculty of Medicine and Pharmacy, 35 Al.I. Cuza st., 800216, Galați, Romania, e-mail: mariana_lupoae@yahoo.com
- ² "Ovidius" University of Constanța, Faculty of Natural and Agricultural Sciences, 124, Mamaia blvd., 900527, Constanța, Romania, e-mail: dcoprean@uoc.ro, Academy of Romanian Scientists 54 SplaiulIndependentei 050094, Bucharest
- ³ Natural Sciences Museum Complex Galați, Botanical Garden, 6A Regiment 11 Siret st., 800340, Galați, Romania, e-mail: paul_lupoae@yahoo.com

Abstract

The purpose of this paper was to evaluate the antioxidant effect of certain extracts obtained from indigenous plants (Allium ursinum and Alliaria petiolata) by introducing them in the diet of the rainbow trout reared in conditions of overpopulation in a recirculating system. At the end of the experimental period, the following investigations were accomplished: biochemical (MDA, TAC, GLU, TP, IgM), hematological (Ht, Hb, MetHb, E, VEM, HEM CHEM) and biotechnological (SGR, FCR). The highest significant values (p<0.05) of MDA, compared to the lots treated with vitamin E and Allium extracts, were registered in the muscles and liver of the fish in the witness lot not treated with extracts (the 2% version). In the liver, the significant increases of TAC, compared to the untreated witness lot, are in LotE (p=0.023<0.05), in LotAllium (p=0.05≤0.05) and in LotAlliaria (p=0.01<0.05). The hemoglobin has values between 7.73-7.76 g/dL in LotAllium and between 7.55-7.62 g/dL in LotAlliaria, both being lower than LotE. The HEM values are comparable among the lots, ranging between 69.92-70.15pg, while the CHEM value is higher in LotMN, compared to LotE, LotAllium and LotAlliaria. The IgM has a minimum value of 88.23mg/dL in LotAlliaria (5%) and a maximum value of 92.03mg/dL in LotAlliaria (2%). In both experimental versions, positive correlations were obtained between the SGR and the average values of TAC in the analyzed lots. The final conclusion is that the vegetal extracts of Allium ursinum and Alliaria petiolata used in our experiments can determine, in certain concentrations, the reduction of the stress induced by overpopulation in Oncorhynchus mykiss.

Abreviations: CHEM – concentration of mean corpuscular hemoglobin, \mathbf{E} – number of erythrocytes, \mathbf{FCR} – Feed Conversion Ratio, \mathbf{GLU} – glucose, \mathbf{Ht} – hematocrit, \mathbf{Hb} – hemoglobin, \mathbf{HEM} – mean corpuscular hemoglobin, \mathbf{Ig} \mathbf{M} – immunoglobulin \mathbf{M} , \mathbf{MDA} – malondialdehyde, \mathbf{MetHb} – methemoglobin, \mathbf{PT} – total protein, \mathbf{SGR} – Specific Growth Rate, \mathbf{TAC} – Total Antioxidant Capacity, \mathbf{VEM} – mean corpuscular volume.

Keywords: vegetal extracts, antioxidant effect, lipid peroxidation, rainbow trout, overpopulation.

Introduction

Currently, the objective of the recirculating systems used in aquaculture is the intensification of the fish production which means, first of all, high population densities in the rearing basins. This aspect favors the maintenance of the culture biomass in conditions of oxidative stress, from the point of view of the ecophysiological comfort. The limited space in which the culture biomass is forced to live constitutes itself a factor that puts a heavy load on the adaptation mechanisms and which may lead to an immunity decline. Also, overpopulation influences the easy transmission and reproduction of specific pathogenic agents, which means the life conditions become dangerously altered.

Generally, the intensive rearing of animals, including of fish, involves large concentrations of individuals in a limited space. In intensive and superintensive aquaculture, there are rearing systems where the fish biomass can exceed 10-20 Kg/m³ of water [18]. The increase of population density (overpopulation) in confined spaces reduces the vital space proportionally. The consequence is the appearance of neurohormonal modifications with direct implications on the biochemical and hematological values.

The importance of population density in fishery technology was emphasized by numerous authors who studied this aspect for different culture species and different production systems [4], [11], [12], [24-25].

The studies accomplished at international level on *Oncorhynchus mykiss* highlighted that this species, compared to other salmonidae, has few demands in terms of environmental conditions, has a good degree of feed assimilation and thus a fast growth rate in captivity. These technological issues represent an important argument for the rearing in superintensive recirculating system with the purpose of producing consumer trout.

Still, if we take into account the ecobiology of this species (the rainbow trout is a territorial fish), it is desired that the population density in the rearing basins should permit the establishment and maintenance of territories. In these conditions, overpopulation leads to stress. To support this hypothesis, it is accepted that fish agglomeration is a factor of chronic stress which may lead to behavioral and physiological modifications [27-28], [30], [43].

The necessity to obtain high fish productions in recirculating systems, corroborated with a reduction of the oxidative stress determined by overpopulation, stimulated the current research. Consequently, despite the increased progress registered over the recent years and the awareness of the complexity of such an approach made our study begin by the testing of vegetal extracts introduced in the feed of the rainbow trout and by the subsequent evaluation *in vivo* of the antioxidant effect through experimental biochemical, hematological and biotechnological analyses.

Material and methods

The research took place in an aquarium-type superintensive recirculating rearing system, in the Laboratory of the Department of Aquaculture, Environmental Science and Land Register within the "Dunarea de Jos" University, Galati. Under the functional aspect the system was particularized to the technological demands for rearing culture fish species in limited spaces. The scheme of the recirculating system is presented in Figure 1.

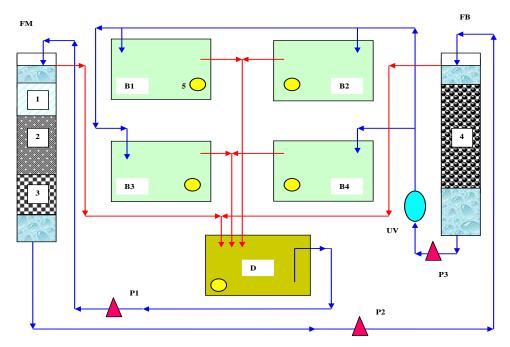


Figure 1. The scheme of the experimental recirculating system Legend: B₁-B₄-rearing units, D -decanting basin, P₁,P₂,P₃-pumps, UV- sterilization lamp, FM- mechanical filter, FB-biological filter, 1-sponge, 2-sand, 3-gravel 4-bactobolts, 5- air nozzle.

The biological material was represented by the one-year-old culture species *Oncorhynchus mykiss*, reared in the same system during the juvenile stage. The biomass used in aquariums, considered admissible, which respected the bearing capacity of the recirculating system, but crucial in the accentuation of the oxidative stress, was 7300±60g /aquarium (73 fish/aquarium with average mass 100±5g/individual). The quantity of feed administered (ration) was 3% of the initial biomass. The feeding frequency was established to twice a day (at 9 a.m. and 5 p.m.). The feed was administered in the form of extruded pellets (Nutra Pro

MP-T), sized 1.7 mm and with the following biochemical composition: crude proteins 50%, crude fats and oils 20%, fibers 10%, crude ash 8%, phosphorus 1.2%, calcium 1%, sodium 0.4%, vitamin A 6000 UI/kg, vitamin D₃ 1200UI/kg. As antioxidant-**known witness-** vitamin E (α -tocopheryl acetate) was used as soft capsules produced by S.C. Biofarm S.A.

Two experimental versions were accomplished. Duration 15 days/version:

- -version a: plant addition 2% in Lot_{Allium} and Lot_{Allium Alliaria}; Lot_{AlliumMN}; Lot_{Allium E}.
- -version b: plant addition 5% in Lot_{Allium} and Lot_{Allium} Allium allium in Lot_{Allium} E. Legend: Lot_{Allium} MN untreated witness lot, Lot_{Allium} E lot treated with vitamin E

Legend: Lot_{Allium MN} – untreated witness lot, Lot_{Allium E} – lot treated with vitamin E (50mg/kg biomass/day).

<u>Preparation of the feed, collection and analysis of the biological samples</u>

The extracts were obtained by double maceration [44] in alcohol 70° for 15 days at room temperature from the two plants – Allium ursinum and Alliaria petiolata (leaves + flowers in equal ratio for each plant) and vitamin E. They were incorporated into the feed after a previous evaporation of the alcohol in Rotavapor by homogenization with gelatin 2%. The drying of the feed was done in the drying oven, at 27°C for 12 hours for the adhesion of the extracts and of vitamin E to the feed surface. Before collecting the biological samples (blood and tissues), the trout individuals were anesthetized. The anesthetic used was a solution of 2phenoxyethanol (1ml for 3 liters of water) in which the fish were immersed for 1 minute. The method we used was collection from the caudal vein in the following stages: tamponing the punctured area with betadine 10% (in order to avoid contaminating the blood with water or mucus), introduction of the syringe needle on the direction located behind the anal fin, puncturing the caudal vein and collection of approx. 1.5-2 ml of blood. The collected blood was poured: a part of it (1ml) in Ependorf tubes with anticoagulant (heparine) for hematological determinations and indicators of oxidative stress, and another part without coagulant for biochemical determinations in other Ependorf tubes.

The harvesting of the internal organs (liver, heart, kidneys, muscles, intestine, gills) was accomplished by dissection, collection and weighing on the analytical balance. The blood samples without heparine were centrifuged for 10 minutes at 3500 rotations/minute and the plasma was separated for the biochemical determinations and for the indicators of the oxidative stress. Of the total amount of blood, 0.1 ml were used immediately for the determination of methemoglobin.

The biochemical determinations were accomplished in the chemistry laboratories within the Faculty of Sciences from "Dunarea de Jos" University, Galati.

The analysis of lipid peroxidation was accomplished by the determination of the concentration of **malondialdehyde** (MDA) and of other substances which

react with the thiobarbituric acid (TBARS), with the help of the method described by Draper and Hadley [7]. The determination of the **total antioxidant capacity** (TAC) from the blood plasma and the tissues of the analyzed samples was done by the method described by Re [31] and Van Den Berg [39]. The quantitative determination of **hemoglobin** was accomplished through the use of the colorimetric method using the Sahli hemoglobinometer and the 0.02 dropper for hemoglobin. This method is based on the fact that hemoglobin mixed with hydrochloric acid turns into hematin hydrochloride. The determination of the **hematocrit** or the volume percentage of erythrocytes, which represents the concentration of erythrocytes in 100 ml of whole blood, was accomplished by the microhematocrit method, which involves centrifuging the blood in capillary tubes (30µL) of heparinized microhematocrit. The determination of **the number of erythrocytes** (red cells) was accomplished according to a classical method using the hemocytometer (the Neubauer counting chamber) and dilution liquid.

The calculation formula used for the determination of the number of erythrocytes, with correction of surface, height and dilution, expressed in millions of cells (e.g. 10⁶/mm³) is as follows:

$$X = \frac{400 \times 200 \times N}{80} = 10000 \times N,$$

where

N-counted erythrocytes in 80 (16x5) squares; **4000**-the volume of squares with 1/20mm side; **200**-blood dilution.

The erythrocyte constants reflect the working degree of the erythrocytes, the content of hemoglobin in each erythrocyte and the ratio between this content and its value. The most important erythrocyte constants are:

The mean corpuscular volume (**VEM**) represents the mean volume of the erythrocyte. The values obtained vary according to age and sex. They are expressed in μ m³ and are calculated according to the formula:

$$VEM = \frac{Ht \times 10}{E}$$

where: Ht is the hematocrit (%); E-is the number of erythrocytes (mill/mm 3) in blood

- The mean corpuscular hemoglobin (**HEM**) represents the mean content of hemoglobin of the erythrocytes. It is expressed in picograms (pg) which represent 10⁻¹² g and are calculated according to the formula:

$$\text{HEM} = \frac{Hb \times 10}{E}$$

where: Hb is the hemoglobin in grams/100 ml of blood. It is the number of erythrocytes expressed in mill./mm³ of blood.

- The concentration in mean corpuscular hemoglobin (**CHEM**) represents the ratio between the corpuscular hemoglobin and its volume, being the most truthful erythrocyte constant. It is expressed in grams/dl of blood and is calculated according to the formula:

$$CHEM = \frac{Hb \times 100}{Ht}$$

where

Hb is the hemoglobin expressed in grams/100 ml of blood;

Ht is the hematocrit (%).

At the end of the experimental period/**version**, the fish were weighed and the bioindicators of technological performance were calculated, namely:

- **The specific growth rate (SGR)** was calculated according to the method described by Hevroy et al. [10]:

SGR=(
$$\ln W_t$$
- $\ln W_0$) x 100 x t⁻¹ (%BW/day), where,

 W_{t} - final biomass; W_{0} -initial biomass; t-experiment duration (days), BW-body weight (g);

- The feed conversion ratio (FCR) was calculated according to the method described by Shalaby et al. [33]:

FCR=
$$Q/S_r$$
, (g/g) , where:

Q-the quantity of feed administered (g); $S_r = W_t - W_0$ (g).

The statistical analysis of the experimental data

The results were expressed as means± and standard deviation (SD). The statistical data were analyzed in the Windows 2007 programs (Microsoft Office Excel, Microsoft Office Word), while the means and standard deviations were calculated according to standard methods for all parameters.

The Student "t" test was used for the comparison of the means. The difference between two means was considered significant for a value of p<0.05.

Results and Discussions

Over the 15 days of experiments, the fish with identification chips from the experimental lots had a slightly agitated behavior, but without pathological symptoms or the presence of parasites on their tegument. We must emphasize that the physic-chemical parameters of the water were maintained in the optimum values for the rearing of this species so as not to create additional stress for them.

If we look at Figure 2, we can see that the level of lipid peroxidation induced by the overpopulation stress is more accentuated for the muscular and liver tissue in the witness lot (a lot not treated with vitamin E or vegetal extracts obtained from *Allium* and *Alliaria*). It seems these tissues are the most sensitive to the overpopulation stress in the rainbow trout.

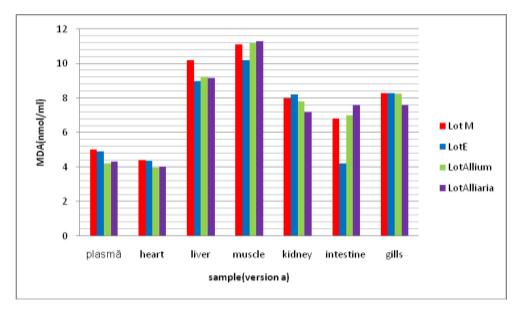


Figure 2. The dynamics of the malondialdehyde index (MDA) from the tissue and blood plasma of fish in version **a** (plant extract 2%)

In what regards the treatment with extracts obtained from *Allium ursinum* and *Alliaria petiolata*, given the overpopulation stress, the reduction of the MAD tissue level is significant (p<0.05) for the liver, both in the 2% concentration and in the 5% concentration of the vegetal extract (see Figure 2 and Figure 3).

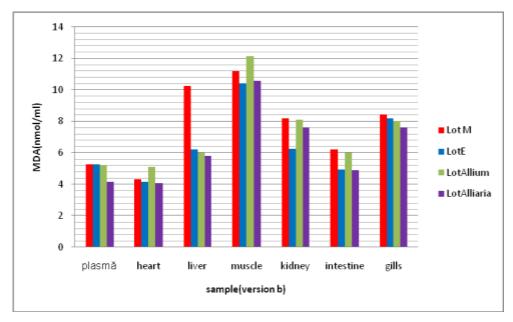


Figure 3. The dynamics of the malondialdehyde index (MDA) from the tissue and blood plasma of fish in version **b** (plant extract 5%)

In the case of the blood plasma, the heart, muscle, intestine, gills and kidneys, the treatment with vegetal extract 2% from both plants (*Allium* and *Alliaria*), given the overpopulation stress, does not reduce the level of lipid peroxidation (expressed by the MAD tissue concentration (see Figures 2 and 3). Vitamin E either does not reduce the level of lipid peroxidation (increased by overpopulation stress) in the same organs.

Similar results were obtained by Metwally [17], who followed the MDA evolution in the crustacean *Litopenaeus vannamei* over a period of 84 days. Indeed, the vegetal extracts introduced in the shrimps diet led to the reduction of the MDA concentration in gills, first insignificantly (at 21 days), and then significantly (at 84 days).

Regarding the total antioxidant capacity, the 2% extract of *Allium* and *Alliaria* increases this capacity, given the overpopulation stress in the rainbow trout, for blood plasma, muscles, and liver. Moreover, the 2% extract of *Alliaria* increases the same antioxidant capacity also in the case of the renal tissue (see Figure 4, version a).

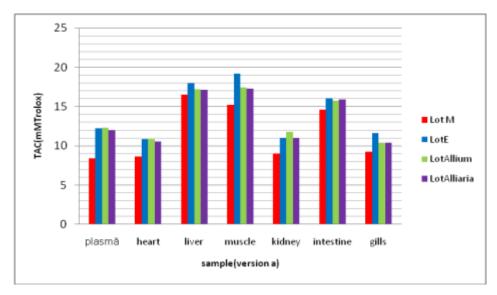


Figure 4. The dynamics of the total antioxidant capacity in tissues and blood plasma of fish in version a (plant extract 2%)

Given the overpopulation stress, the treatment was done with the 5% extract of *Alliaria petiolata*, and there was an increase of the total antioxidant capacity in the case of the blood plasma, heart, liver and muscle as well, compared to the untreated lot (see Figure 5).

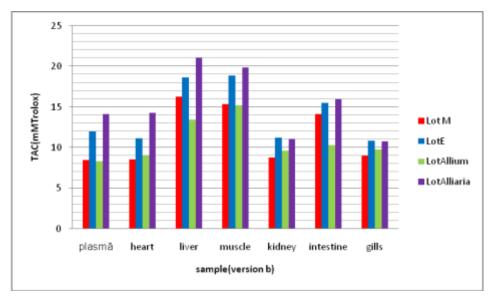


Figure 5. The dynamics of the total antioxidant capacity in tissues and blood plasma of fish in version b (plant extract 5%)

Given the overpopulation stress, vitamin E determined an increase of the total antioxidant capacity in plasma, liver and muscles (experimental version a), or in liver and intestine (experimental version b; see Figure 5). If we trace an overview of the experimental results where we followed the effect of the 2% and 5% vegetal extracts from *Allium* and *Alliaria* in the rainbow trout subjected to overpopulation stress, we observe that the vegetal extracts were at least equal to vitamin E in terms of stimulating the total antioxidant capacity (see Figures 4 and 5).

These results allow us to appreciate the antioxidant effect of the used extracts, which can be attributed to the content in bioactive compounds such as: flavonoids, volatile oils, vitamins and others. In support of this discovery, some authors attributed to vitamin C an indirect antioxidant effect [21], [37], while others signaled a positive, modulating effect of vitamins C and E in response to stress [9], [13], [22], [32], [35].

For a more complete evaluation of the tested vegetal extracts in the attenuation of the induced oxidative stress, in the present experiment we also followed the evolution of certain hematological parameters. In this regard, for the appreciation of the physiological state of the fish, we accomplished the hemoglobin dosing, considered a precise and quick test for the checking of the hematological homeostasis [18]. Hemoglobin is the respiratory pigment with important role in the increase of the blood's capacity to transport oxygen. In our experiment (see Table 1), we observed that vitamin E, administered in conditions of overpopulations stress in the rainbow trout, determines modifications of the hemoglobin content and number of erythrocytes (comparison between Lot_E and Lot_{MN} ; MN = untreated witness).

Table 1

Variation of the hematological parameters in fish at the end of the experiment

Hematologic	Lot _{MN}	Lot _E	Lot _{Allium}		Lot _{Alliaria}	
al			2%	5%	2%	5%
parameter						
Ht (%)	41±5.23 ^b	40±3.38 ^b	36±6.22 ^b	32±6.99 ^b	36±3.89 ^b	38±5.35 ^b
Hb(g/dL)	6.72±0.10 ^{ab}	7.80±0.08 ^a	7.76±0.07 ^a	7.73±0.89 ^b	7.62±0.58 ^b	7.55±0.68 ^b
No.E x10 ⁶ /mm ³	1.05±0.03 ^{ab}	1.22±0.04 ^b	1.28±0.02 ^a	1.12±0.11 ^b	1.29±0.03 ^a	1.27±0.11 ^b
VEM(μm³)	365.10±12.5 ^b	420.33±10.6 ^b	385.17±24.0 ^b	386.55±15.2 ^b	419.31±10.7 ^b	421.04±20.1 ^b
HEM(pg)	70.15±4.48	70.02±3.07	69.95±7.29	69.90±9.02	69.97±4.80	69.92±5.83
CHEM g/dL	18.46±2.0 ^b	18.50±1.03 ^b	18.90±1.92 ^b	18.58±2.11 ^b	18.58±3.41 ^b	18.49±1.47 ^b

The various superscript letters in the columns indicate significant(a) and insignificant(b) differences with cu p<0.05; the statistical analysis was accomplished on the horizontal row thus: Lot_{MN} compared to Lot_E and the lots not treated with the extracts.

The vegetal extracts of *Allium ursinum* and *Alliaria petiolata* influence the level of the hematological parameters. Thus, the *Allium* extract in 2% concentration determines the increase of hemoglobin and the number of erythrocytes, while the *Alliaria* extract in 2% concentration only increases the number of erythrocytes (see Table 1). The increase in the number of erythrocytes, and thus of the quantity of hemoglobin in the lots treated with *Allium* and *Alliaria* extracts could be the result of the direct action of the active principles from these extracts on the level of production of the figurate elements. As we have shown in the experiment where the stress was induced by intoxication with sodium nitrite, any state of stress is associated with more intense energy consumption at the level of cells and tissues, because only this way the cells and tissues can efficiently resist metabolically to the state in which they are.

So, our analyses emphasize the increases of hemoglobin in the lot with vitamin E addition, but also in the lots with addition of vegetal extracts. This means that a larger quantity of oxygen transported to the tissues where the intensified oxidative metabolic processes and production of energy ensure their increased resistance to stress. In some research, it is shown that the significant reduction of the amount of hemoglobin in blood can affect the amount of oxygen for tissues and the result is the slowing down of the metabolic rate, thus a reduced production of energy [1]. Also, the significant reduction of the amount of hemoglobin can be caused by the increase of the rate of hemoglobin destruction, or the reduction of the rate of its synthesis, given the state of stress. According to other authors [23], these hemoglobin modifications are determined by different forms of stress which in turn determine the rapid increase of the hemoglobin concentration due to the recruiting of erythrocytes from the spleen and the hemoconcentration installed as a result of the loss of plasmatic water.

The hematocrit (the volume percentage of erythrocytes) does not modify significantly in the lot with addition of vitamin E, as in the lots treated with extracts of *Allium ursinum* and *Alliaria petiolata*, compared to the witness lot (Lot_{MN}). According to some research [23], the increase of hematocrit in fish is joined by an increase of blood viscosity, which is considered the superior limit of the strategy of adaptation to stress. Other authors [15], [42] remarked the influence of different stress factors on the hematocrit in certain culture species, which determined a dynamics of the hematocrit in a very wide range, between 20-40%.

The slight decrease of the hematocrit joined by an increase of the hemoglobin influenced the increase in the number of erythrocytes in Lot E, Lot_{Allium} and Lot_{Alliaria}. The data in the specialized literature account for the fact that in *Oncorhynchus mykiss* the number of erythrocytes can vary from 0.85 to 1.50(x10⁶/mm³) depending on age, sex, physiological state etc. The values obtained by us fit in the superior limit, with the highest value in Lot_{Alliaria} (2%)

and Lot_{Allium} (2%). These values are significantly increased (p<0.05), compared to the untreated witness lot.

Similar results were obtained by Valenzuela et al. [38] in a study regarding the effect of physical stress induced by the increase of temperature and continuous application of light, which led to the increase of hemoglobin concentration and number of erythrocytes.

The analysis of the erythrocyte constants (VEM, HEM, CHEM) can help to detect the presence of physiological ailments in the process of hemoglobin formation and offers information regarding the size, shape and hemoglobin load of erythrocytes.

Under the influence of the vegetal extracts and vitamin E, the constant erythrocyte values from the analyzed samples registered the following modifications:

- The mean corpuscular volume (VEM) increased insignificantly (p>0.05) in the analyzed lots compared to the untreated witness lot;
- The mean corpuscular hemoglobin (HEM) registered relatively constant values, compared to Lot_{MN};
- The concentration of mean corpuscular hemoglobin (CHEM) increased insignificantly (p>0.05), compared to the untreated witness lot.

The analysis of the values of the three erythrocyte constants allow us to formulate the observation that the VEM, HEM and CHEM fit in the limits considered optimal for the rearing of salmonidae. However, we remark that the values in the inferior limit are found in the witness lot. Also, the values of the erythrocyte constants in the lots fed with vegetal extracts are comparable with those of the lot treated with vitamin E.

The present study continued with the analysis of biochemical parameters of blood: glycemia, total plasma proteins and plasma immunoglobulin M. In the case of our experiment, where the stress was induced by overpopulation in the rainbow trout, we observed that neither the vegetal extracts of *Allium ursinum* and *Alliaria petiolata*, nor vitamin E modify significantly the glycemia level compared to Lot_{MN}.

In other words, we can say that in conditions of overpopulation stress, neither the vegetal extracts of *Allium ursinum* and *Alliaria petiolata*, nor vitamin E enhance the mechanisms of glucose release from storage in order to be consumed by the peripheral tissues. It is possible that the overpopulation stress should be easier to bear by the fish organism, compared to the stress induced by intoxication with sodium nitrite (unpublished personal data), as the fish cope more easily with this type of stress without additional energy consumption (see Table 2).

The plasma sugars represented especially by glucose are normally found at values between 40-90mg/100mL in the fish blood, while in the healthy rainbow trout, they range around 71mg/100mL of blood [12]. According to some authors,

glycemia can be influenced by a number of technological factors and constitutes an important marker for the stress states in fish. The manipulation stress can lead to a significant increase of the glycemia up to values of 137mg/dL [2]. Other authors state that an increase in glycemia represents the response of the organism in fish exposed to acute or chronic stress [19-20], [41], [3], [29], [36].

Table 2
Biochemical parameters in fish in conditions of overpopulation

Sample	Statistical indexes	GLU mg/dL	TP g/dL	Ig M mg/dL		
Lot _{MN}	X±ES	120.2±10.01	3.42±0.22	90.62±4.06		
	n	4	4	4		
	X±ES	112.0±8.48	4.30±0.08	89.00±3.22		
Lot_{E}	n	4	4	4		
	±M%	-6.82	+25.73	-1.79		
	plant concentration 2%					
	X±ES	87.20±3.19	4.25±0.23	90.1±1.44		
	n	5	4	5		
Lot _{Allium}	±M%	-27.45	+24.27	-0.57		
	plant concentration 5%					
	X±ES	89.13±6.03	3.15±0.31	91.6±0.2		
	n	5	5	5		
	±M%	-25.84	-7.9	+1.08		
	plant concentration 2%					
	X±ES	87.77±7.22	3.60±0.87	92.03±5.07		
	n	5	5	5		
Lot _{Alliaria}	±M%	-26.98	+5.26	+1.5		
	plant concentration 5%					
	X±ES	85.94±5.80	4.46±0.44	88.23±1.20		
	n	4	5	5		
	±M%	-28.50	+30.41	-2.6		

 $\begin{tabular}{ll} \textbf{Legend:} Lot_M-witness\ lot,\ Lot_E\ (addition\ of\ vitamin\ E),\ Lot_{Allium}\ (plant\ addition),\ Lot_{Allium}\ (pl$

If we refer to the plasma proteins and immunoglobulin M, the results of our experiments show that neither the vegetal extracts of *Allium ursinum* or *Alliaria petiolata*, nor vitamin E modify significantly this blood parameter in a stress state induced by overpopulation. At least from the point of view of the biochemical parameters studied, we can say that stress induced by intoxication with sodium nitrite (unpublished personal results) is more difficult to bear by the organism of the rainbow trout compared to the stress induced by overpopulation.

According to some authors [34], the normal values of the total serum proteins in fish range between 3.5 and 5.5g/dL. In *Oncorhynchus mikiss*, the optimal values published in specialty articles are different: 3.64-3.80 g/dL [14]; 2.52-3.16 g/dL [16]; 2.92 g/dL [40]; 3.7-4,7 g/dL [5]. Still, these can display ample variations according to species, age, sex, water temperature, season, quality and quantity of feed [26].

The proteinemy in our samples is slightly increased compared to the lot not treated with vegetal extracts with percentage rises between 5.26-30.41%, with one exception in Lot_{Allium} (5% extracts concentration), where the protein level decreases by 7.9%. The explanation we propose is that the variations of plasma proteins may be due to the intensification of the appetite manifested in the fish in Lot_{Alliaria}, Lot_E and Lot_{Allium} (2%). This phenomenon may be due to the increased protein content in the feed (50% crude protein), associated with the intake of antioxidants in the vegetal extracts tested. The 5% *Allium ursinum* concentration from the extracts used proved to be less agreeable and thus less assimilated by the fish. The increase of the protein level in blood was also signaled by Farahi et al. [8], who added garlic (*Allium sativum*) in the fish feed.

In the ontogenetic development, the first class of immunoglobulin which appears is the immunoglobulin M class (IgM) and they represent the antibodies that appear after the first contact with the antigen. The IgM values can be influenced by a number of exogenous and endogenous factors and range between 1-100mg/dL [26]. The values obtained by us register both increases and decreases compared to the witness lot not treated with vegetal extracts. The most considerable percentage decrease (2.6%) was registered in Lot_{Alliaria} (5%), while the increases were of +1.8% la Lot_{Allium} (5%). All the values fit in the superior limit of the optimum range, without significant statistical differences. Most likely, the stress induced by overpopulation triggers the organism's reaction to this type of stress, but within the normal values.

The study of growth performance by means of the main biotechnological indicators (see Table 3) represents a way to appreciate quantitatively the normality degree of the physiological state of the fish organism. This aspect allowed us to evaluate the dynamics of the culture biomass by the analysis of the specific growth rate (SGR) and of the feed conversion factor (FCR). The addition of vitamin E in the fish feed led to an increase of the SGR by 42.5% compared to the witness lot not treated with vegetal extracts. The vegetal extracts added to the feed were tolerated by the fish differently, a fact which was noticed both during the experiment and after the analysis of the first samples. The highest percentage increases of the SGR, compared to Lot_{MN}, were registered in Lot_{Alliaria} (5%), comparable to Lot_{Alliaria} (2%).

Similar results were obtained in other research as well [8], such as different amounts of *Allium sativum* being introduced in the diet of the species *Oncorhynchus mykiss*, or common sea-buckthorn in the feed of carp [6].

Table 3
Biotechnological indicators in the rainbow trout in conditions of overpopulation

Biotechnological indicators		Lot _{MN}	Lot _E	Lot _{Allium}	Lot _{Alliaria}				
version a plant concentration 2%									
FCR (g/g)		2.28	1.54	1.52	1.44				
SGR	%BW/day	1.2	1.71	1.73	1.81				
	%	100	142.5	144.17	150.83				
version b plant concentration 5%									
FCR		2.31	1.85	2.19	1.41				
(g/g)									
SGR	%BW/day	1.19	1.45	1.24	1.84				
	%	100	121.84	104.2	154.62				

Legend: FCR- Coefficient of feed conversion; SGR- Specific growth rate

If we take into account the specific growth rate, corroborated with the feed conversion factor and the 98.75% level of fish survival, our results are optimistic. Our study shows that the feed supplementation with vegetal extracts leads to the increase of the total antioxidant capacity and the decrease of the lipid peroxidation level. These changes of the antioxidant status are positively correlated with the specific growth rate (see Figure 6).

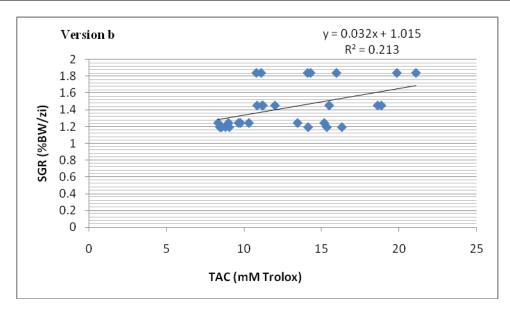


Figure 6. Correlation between the specific growth rate (SGR) and the total antioxidant capacity (extract concentration 5%, n=28)

Based on the experimental data obtained, we can state that the addition of vegetal extracts from *Allium ursinum* and *Alliaria petiolata* in the feed helps to maintain the balance between oxidants and antioxidants, preventing membrane deteriorations caused by oxidative stress.

Conclusions

- 1. The *Allium ursinum* and *Alliaria petiolata* extracts have antioxidant properties in certain concentrations;
- 2. The antioxidant properties of the vegetal extracts used are comparable to those of vitamin E (used as known antioxidant);
- 3. The intensity of the stress induced by various free radicals, including those of oxygen, appreciated according to the malondialdehyde (MDA) value, is different in the analyzed organs;
- 4. Overpopulation triggers an increase of lipid peroxidation, especially in liver and muscles:
- 5. The tissue level of malondialdehyde depends on the nature of the extract. The vegetal extracts from both species are comparable as antioxidant effect in **version a** (concentration 2%);
- 6. The *Alliaria petiolata* extract (concentration 5%) influences positively the total antioxidant capacity, compared to the untreated witness lot (standard feed) and with Lot_E (addition of vitamin E);

- 7. The erythrocyte fragility is correlated with the intensity of stress induced by free radicals, including oxygen free radicals, in the lot not treated with vegetal extracts or vitamin E (considered by us witness lot);
- 8. The tested vegetal extracts favor the increase of hemoglobin level in blood, within physiological limits;
- 9. It was observed that neither the vegetal extracts from *Allium ursinum* and *Alliaria petiolata* nor vitamin E modify significantly the glycemia level compared with the untreated witness lot;
- 10. What results from the analysis of the biotechnological indicators is an active feeding of the fish in case of **version a** (2% extracts) and differentiated in the case of **version b** (5% extracts).
- 11. Consequently, after the installation of oxidative stress by overpopulation, it was observed that the treatment with extracts from *Allium ursinum* and *Alliaria petiolata*, leads to an attenuation of the level of lipid peroxidation and to an enhancement of the total antioxidant capacity.

References

- [1]. Atamanalp M., Yanyk T., Halilo Ulu I., Aras M.S., Alterations in the hematological parameters of rainbow trout (*Oncorhynchus mykiss*) exposed to cypermethrin. Israeli , Journal Aquaculture Bamidgeh 54, 2002, 99-103.
- [2]. Barry T. P., Lapp A. F., Kayes T. B., Malison J. A., Validation of a microtitre plate ELISA for measuring cortisol in fish and comparison of stress response of rainbow trout (*Oncorhynchus mykiss*) and lake trout (*Salvelinus namaychus*), Aquaculture, 117, 1993, 351 363.
- [3]. Biron M., Benfey T.J., Cortisol, glucose and haematocrit changes during acute stress, cohort sampling, and the diel cycle in diploid and triploid brook trout (*Salvelinus fontinalis* Mitchill). Fish Physiol. Biochem. 13, 1994, 153–160.
- [4]. Carr B.A., Aldrich D.V., Population density effects on the behavior and feeding of young striped mullet (*Mugil cephalus*) in 37,8 liter aquaria, Journal World Maricult. Soc 13, 1982, 254-260.
- [5]. Ceschia G., Ciorgeti G.M., Lo Greco P., Parametri ematici ed attivita enzimatica nella trota iridea (*Salmo gairdneri*) allevata. Arch. Vet. Italia 29: 1978, 141-144.
- [6]. Csep L., Bud I., Chirila F., Disease resistance effect of sea-buckthorn (*Hippophae rhamnoides* L.) added in the fish diet, AACL Bioflux 3(5), 2010, 339-346.
- [7]. Draper H.H, Hadley M., Malondialdehyde determination as index of lipid peroxidation. Methods. Enzymol. 186:1990,421-431.
- [8]. Farahi A., Milad K., Mohammad S., Mohsen S.I., Morteza D.S., Effect of garlic (*Allium sativum*) on growth factors, some hematological parameters and body compositions in rainbow trout (*Oncorhynchus mykiss*), AACL Bioflux, 3(4), 2010, 317-323.

- [9]. Henrique MMF., Gomes EF., Gouillou-Coustans M.F., Oliva-Teles A., DAVIES S.J., Influence of supplementation of practical diets with vitamin C on growth and response to hypoxic stress of seabream, *Sparus aurata*. Aquaculture161, 1998, 415-426.
- [10]. Hevroy E.M., Espe M., Waagbo R., Sandness K., Rund M.M., Hemre G., Nutrition utilization in Atlantic salmon (*Salmo salar*) fed increased level of fish protein hydrolyses during a period of fast growth, Aquaculture Nutrition 11, 2005, 301-313.
- [11]. Jorgensen E.H., Christiansen J.S., Jobling M., Effects of stocking density on food intake, growth performance and oxygen consumption in Arctic charr (*Salvelinus alpines*), Aquaculture 110, 1998, 191-204.
- [12]. Kebus M. J., Effects of Rearing Density on the Stress Response and Growth of Rainbow Trout Journal of Aquatic Animal Health, 4: 1992,1-6.
- [13]. Kolkovski S., Czesny S., Yackey C., Moreau R., Cihla F., Mahan D., Dabrowski K., The effect of vitamins C and E in (n=3) highly unsaturated fatty acids-enriched *Artemia nauplii* on growth, survival, *Stizosteidon vitreum* larvae , Aquaculture Nutr., 6, 2000, 199-206.
- [14]. Lupi Paola, Vigiani V., Mecatti M., Contribution to the definition of the metabolic profile of faemed rainbow trout (*Oncorhynchus mykiss*), Italian Journal Animal Science, 5, 2006, 63-71.
- [15]. Martinez FJ., Garcia-Riera MP., Canteras M., Costa J., Zamora S., Blood parameters in rainbow trout (*Oncorhynchus mykiss*): simultaneous influence of various factors, Comp. Biochem. Physiol. 107 A: 1994, 95-100.
- [16]. Melotti P., Roncarati A., Angelloti L., Dees A., Magi GE., Mazzini C., Bianchi C., Casciano R., Effects of rearing density on rainbow trout welfare, determined by plasmatic and tissue parameters, Italia Journal Animal Science, 3: 2004, 393-400.
- [17]. Metwally M.A.A., Effects of garlic (*Allium sativum*) on some antioxidant activities in Tilapia Nilotica (*Oreochromis niloticus*), World Journal of Fish and Marine Science 1 (1): 2009, 56-64.
- [18]. Misăilă Elena, Cercetari biochimice si ecofiziologice asupra profilului metabolic la unii pești de cultură (Teză de doctorat), Universitatea Al. I. Cuza Iași, 1998.
- [19]. Morales A.E., Garcia-Rejon L., De la Higuera M., Influence of handling and/or anaesthesia on stress response in rainbow trout. Effects on liver primary metabolism. Comp. Biochem. Physiol.95A, 1990, 87–93.
- [20]. Morales A.E., Cardenete G., Abellan E., Garcia-Rejon L., Stress-related physiological responses to handling in the common dentex (*Dentex dentex* Linnaeus, 1758), Aquac. Res. 36, 2005, 33–40.
- [21]. Moreno JJ., Mitjavila MT., The degree of unsaturation of dietary fatty acids and the development of atherosclerosis (Review), J. Nut. Biochem. 14, 2003, 182-195.
- [22]. Montero D., Kalinowski T., Obach A., Robaina L., Tort L., Caballero M.J., Izquerdo M.S., Vegetable lipid sources for gilthead seabream (*Sparus aurata*): effects on fish health. Aquaculture 225, 2003, 353-370.
- [23]. Nicula M., Fiziologia organismelor acvatice I., Fiziologia peştilor, I, Editura Mirton Timişoara, 2004.

- [24]. Papoutsoglou SE., Papaparaskeva-Papoutsoglou, Alexis MN., Effect of density on growth rate and production of rainbow trout (*Salmo gairdneri* Rich.) over a full rearing period. Aquaculture 66: 1987, 9-17.
- [25]. Papoutsoglou SE., Voutsinos G.A., Panetsos F., The effect of photoperiod and density on growth rate of *Oreochromis aureus* (Steindachner) reared in a closed water system. Anim. Sci. Rev. 11, 1990, 73-87.
 - [26]. Patriche Tanti, Imunitatea la pești, Editura didactică și pedagogică R.A., 2008.
- [27]. Pickering A.D., Rainbow trout husbandry: management of the stress response, Aquaculture 100, 1992,125–139.
 - [28]. Pickering A.D., Growth and stress in fish production. Aquaculture 111, 1993, 51–63.
- [29]. Pottinger T.G., Carrick T.R., A comparison of plasma glucose and plasma cortisol as selection markers for high and low stress responsiveness in female rainbow trout, Aquaculture 175, 1999, 351–363.
- [30]. Puangkaew J., Kiron V., Satoh S., Watanabe T., Antioxidant defense of rainbow trout (*Oncorhynchus mykiss*) in relation to dietary n-3 highly unsaturated fatty acids and vitamin E contents. Comp. Biochem. Physiol. C 140, 2005, 187–196.
- [31]. Re R., Pellegrini R., Proteggente A., Pannala A., Yang M., Rice-Evans C., Antioxidant activity applying an improved ABTS radical cation decolorization assay, Free Radical Biology and Medicine, 26: 1999, 1231-1237.
- [32]. Sakakura Y., Koshio S., Lida Y., Tsukamoto K., Kida T., Blom JH., Dietary vitamin C improves the quality of yellowtail (*Seriola quinqueradiata*) seedlings. Aquaculture 161, 1998, 427-436.
- [33]. Shalaby A.M., Khattab Y. A., Abdel Rahman A.M., Effect of garlic (*Allium sativum*) and chloramphenicol on growth performance, physiological parameters and survival of *Nile tilapia* (*Oreochromis niloticus*), Journal Venom Anim Toxins incl Trop Dis 12(2), 2006, 172-201.
- [34]. Swicki AK, Douglas P., Fish diseases diagnosis and prevention methods, FAO Project International Workshop and training course in Poland, 1993, 105-111.
- [35]. Tago A., Yamamoto Y., Teshima S., Kanazawa A., Effects of 1,2-di-20:5-phosphatidylcholine(PC) and 1,2-di-22:6-PC on growth and stress tolerance of Japanese flounder (*Paralichthis olivaceus*) larvae. Aquaculture 179, 1999, 231-239.
- [36]. Trenzado C.E., Carrick T.R., Pottinger T.G., Divergence of endocrine and metabolic responses to stress in two rainbow trout lines selected for differing cortisol responsiveness to stress. Gen. Comp. Endocrinol. 133, 2003, 332–340.
- [37]. Upston J.M., Terentis A.C., Stocher R., Tocopherol-mediated peroxidation of lipoproteins: implications for vitamin E as a potential antiatherogenic supplement, FASEB Journal 13, 1999, 977-994.
- [38]. Valenzuela A.E., Silva V.M., Klempau A.E., Effects of different artificial photoperiods and temperatures on haematological parameters of rainbow trout (*Oncorhynchus mikiss*), Fish physiology biochemistry 34; 2008, 159-167.
- [39]. Van der Berg R., Haenen G.R., Van den Berg H., Bast A., Applicability of an improved Trolox equivalent antioxidant capacity (TEAC) assay for evaluation of antioxidant capacity measurement of mixtures, Food Chemistry, 1999, 66:511-517.

Mariana LUPOAE, Dragomir COPREAN, Paul LUPOAE

- [40]. Velisek J., Svodobova Z., Anaesthesia of rainbow trout (*Oncorhynchus mykiss*) with 2-phnoxyethanol: acute toxicity and biochemical blood profile, Acta Vet. ,2004, 73(3): 379-384.
- [41]. Vijayan M.M., Moon T.W., Acute handling stress alters hepatic glycogenmetabolismin food-deprived rainbowtrout (*Oncorhynchus mykiss*), Can. Journal Fish. Aquat. Sci. 49, 1992, 2260–2266.
- [42]. Vosyliene M.Z., Kazlauskiene N., Alterations in fish health state parameters after exposure to different stressors, Acta Yoologica Lituanica, Hydrobiologia, 1999,9(2): 83-94.
- [43]. Wedemeyer G.A., Physiology of fish in intensive culture systems, Northwest Biological Science Center. National Biological Service. U.S. Department of the Interior. Chapman & Hall. International Thompson Publishing, 1996.
 - [44]. ***FARMACOPEEA ROMÂNĂ, Ediția a X-a, Editura Medicală București, 1993.